



A highly efficient strategy for enhancing the adsorptive and magnetic capabilities of biochar using Fenton oxidation



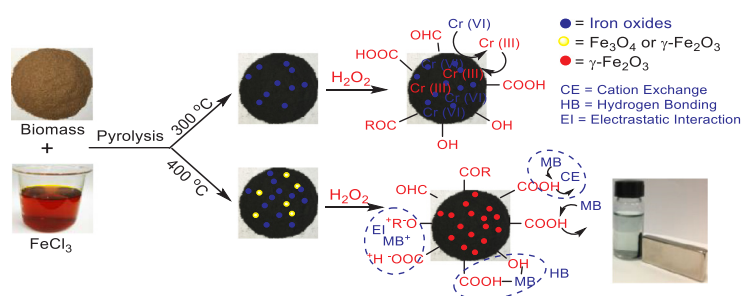
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GRAPHICAL ABSTRACT



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ABSTRACT

Fenton modification, involving iron-promoted pyrolysis followed by H₂O₂ oxidation, was first employed to improve the adsorptive and magnetic capabilities of biochar. Modified biochars were prepared from rubber tree bark and coconut shell through iron-promoted pyrolysis and subsequent H₂O₂ oxidation, and their adsorption behaviors toward Cr (VI) and MB were evaluated in aqueous solution. The modified biochars pyrolyzed at 300 and 400 °C displayed much higher adsorption capabilities than corresponding pristine biochars for Cr (VI) and MB, respectively, ascribing to introduction of COOH, C=O and C–O groups by Fenton oxidation. More importantly, saturation magnetization could be enhanced by transforming nonmagnetic iron oxides into γ-Fe₂O₃ through H₂O₂ oxidation. The removal of Cr (VI) and MB could be primarily contributed to the adsorption of biochar matrix by reduction/hydrogen bonding/cation exchange/electrostatic interaction and hydrogen bonding/cation exchange/electrostatic interaction, respectively. This would provide a novel and efficient strategy for making highly adsorptive magnetic biochar.

1. Introduction

Magnetic modification is an attractive strategy for overcoming the difficulty in solid–liquid separation of biochar, provides great advantages for water treatment using biochar (Wang et al., 2019; Yi et al.,

2020). Mostly, it is realized by introducing magnetic medium through either pyrolysis (Han et al., 2016; Trakal et al., 2016) or post-coating (Mohan et al., 2014a; Wang et al., 2017) using low-cost and less toxic iron salts as precursor. One common method is zero-valent iron (nZVI) doped modification, in which ZVI is generated on biochar matrix

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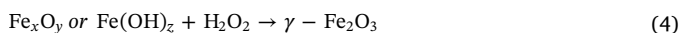
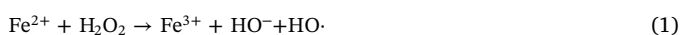
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through reduction of iron species by NaBH₄ (Wang et al., 2017; Yang et al., 2018). The modification can lead to the increase of adsorption capability due to the complementation of ZVI and biochar matrix (Devi & Saroha, 2014). Comparing to ZVI doped modification, magnetic iron oxide-doped modification should be more attractive for practical application as producing oxidatively stable Fe₃O₄- and/or γ-Fe₂O₃-doped biochar (Mohan et al., 2014b; Zhang et al., 2013). Great advance has been made in this magnetic modification in the past decade, but some defects such as generation of nonmagnetic iron species (Reguyal et al., 2017) and loss of active adsorption sites (Karunanayake et al., 2018; Reguyal et al., 2017) still remain to be resolved.

As generating oxygen-containing groups which have good affinity to heavy metals and organic molecules (Huff & Lee, 2016; Mandal et al., 2017), oxidation is considered to be a promising technology for enhancing the adsorption capability of biochar. Especially, H₂O₂ oxidation, which can tune the physiochemical properties of biochar by introducing oxygen containing groups (Huff & Lee, 2016) as well as modulating morphology and pore structure (Tan et al., 2011), is attractive for environment remediation as producing less harmful residues. In last decades, many H₂O₂ modifications were developed to engineer different nonmagnetic biochars for removal of heavy metals such as Cu²⁺, Pd²⁺, Cd²⁺ and Ni in aqueous solution (Wang et al., 2018; Xue et al., 2012; Zuo et al., 2016). In addition, H₂O₂-pretreatment and microwave activation were combined to enhance the adsorption capability of Prosopis juliflora biochar for remazol brilliant blue and methylene blue (Nair & Vinu, 2016). However, high concentration of H₂O₂ was usually required for these modifications and low efficiency was obtained for improving adsorption capability of biochar due to the weak oxidizing ability of H₂O₂, seriously restricting wide application of H₂O₂ modification.

Fenton oxidation, an iron-catalyzed advanced oxidation, can rapidly generate highly active HO· radical to oxidize different functional groups into diversified oxygen-containing groups (Zhu et al., 2019). The combination of iron modification and H₂O₂ modification (Fenton modification) should complement each other for regulating the physiochemical properties of biochar. H₂O₂ oxidation of biochar matrix can be promoted by iron significantly, generating a large number of oxygen-containing groups including OH, COOH and C=O on biochar (eqs (1), (2)). Meanwhile, H₂O₂ decomposition (eq (3)) can also be accelerated to massively produce O₂ for modulating the morphology of biochar. On the other hand, nonmagnetic iron species such as ferrous oxides and ferric hydroxides should be transformed into γ-Fe₂O₃ with oxidation and dehydration of H₂O₂ (eq (4)), benefiting to enhancing saturation magnetization as well as increasing active adsorption sites (Zhang et al., 2020). Hence, Fenton modification can multi-directionally regulate the physiochemical properties of biochar, providing a promising strategy for addressing the drawbacks in both magnetic iron oxide-doped modification and H₂O₂ modification simultaneously. Nevertheless, Fenton oxidation has been less explored for precisely engineering biochar for efficient removal of pollutants by now.



Dye and heavy metal pollution has posed a serious threat to human and ecological environment. Hexavalent chromium (Cr (VI)) and methylene blue (MB) are common contaminants in aquatic environments by industrialization such as mining, electroplating, textile and dyeing (Qada et al., 2006; Pradhan et al., 2017). As a low-cost and renewable adsorbent, biochar has been widely applied to remove Cr (VI) (Chen et al., 2015; Nasuha & Hameed, 2011; Zhao et al., 2018) and MB (Daneshvar et al., 2017; Zhang & Gao, 2013) from aqueous solution in the past decade. Importantly, considerable progress has already been

made in preparation of magnetic biochar for treating Cr (VI)-contaminated water due to the advantage of easy separation. For example, ZVI/biochar composites were prepared for adsorbing Cr (VI) with relatively low adsorption capabilities (Zhou et al., 2014), where the biochars mainly acted as supporter and electron-transfer medium. Multi-step synthesized Fe₃O₄- and γ-Fe₂O₃-doped magnetic biochars were demonstrated to exhibit moderate and high adsorption capabilities toward Cr (VI) (Han et al., 2016; Shi et al., 2018), respectively. Recently, surface functionalization was developed for enhancing the adsorption capabilities of Fe₃O₄/biochar and γ-Fe₂O₃/biochar composites (Mian et al., 2018; Yin et al., 2020), confirming biochar matrix could contribute greatly to the adsorption capability of magnetic biochar. By contrast, despite ZVI- and iron oxide-doped magnetic biochars were also made for adsorbing MB, inferior adsorption capabilities were obtained (Mubarak et al., 2015; Nasuha & Hameed, 2011).

Herein, Fenton oxidation was employed to engineer two biochars made from bio-wastes rubber tree bark and coconut shell for removing Cr (VI) and MB in aqueous solution. For comparison, the corresponding H₂O₂- and FeCl₃-modified biochars were also prepared and their adsorption capabilities were estimated. The objectives of this study were to: 1) establish a general method for improving the adsorption capability of biochar toward Cr (VI) and MB using Fenton oxidation; 2) evaluate Fenton oxidation for enhancing saturation magnetization of magnetic biochar; 3) illuminate the adsorption mechanism of the modified biochars toward Cr (VI) and MB.

2. Materials and methods

2.1. Materials

The rubber tree bark and coconut shell were collected from a farm around Danzhou city in Hainan province of China. The chemicals including HCl, KOH, H₂SO₄, FeCl₃, H₂O₂, MB and K₂Cr₂O₇ were analytical reagent grade and purchased from Aladdin and Macklin. Different concentrations of Cr (VI) and MB solutions were prepared by dissolving K₂Cr₂O₇ and MB in deionized water (water treatment system, LD-UPW-20, Leading, Shanghai, China), respectively.

2.2. Preparation of biochars

The rubber tree bark was crushed into powder (60 mesh). The powder (10 g) and 2 mol/L FeCl₃ solution (80 mL) were mixed and magnetically stirred at room temperature for 2 h, then aged at 70 °C for 30 min. The mixture was filtered to get iron-loaded biomass, and dried at 60 °C for 48 h. The dried sample was ground and pyrolyzed at the target temperature (200 °C, 300 °C, 400 °C, 500 °C, 600 °C) for 1 h with 5 °C/min heating rate under nitrogen. After cooling to room temperature, the sample was washed and dried at 60 °C to get Fe-modified biochar. The biochars were named as Fe200, Fe300, Fe400, Fe500 and Fe600 based on pyrolysis temperature, respectively. The Fe-modified biochar (3 g) was treated with H₂O₂ solution (60 mL) for 2 h, washed and dried at 60 °C to obtain Fenton-modified biochar. The Fenton-modified biochars were named as Fe-H₂O₂200, Fe-H₂O₂300, Fe-H₂O₂400, Fe-H₂O₂500 and Fe-H₂O₂600 according to pyrolysis temperature, respectively. The pristine biochar was prepared through pyrolysis of rubber tree bark powder directly at different temperatures (200 °C, 300 °C, 400 °C, 500 °C, 600 °C) and called P200, P300, P400, P500 and P600, respectively. The P300 and P400 were oxidized with H₂O₂ to make H₂O₂-modified biochars (H₂O₂300 and H₂O₂400). They were also modified by reacting with FeCl₃ (2 M) and H₂O₂ in 3/7/60 (g/mL/mL) ratio to make direct Fenton oxidative biochars (Marked as P300-DF and P400-DF). Besides, pristine coconut shell biochars were prepared at 300 and 400 °C (called CP300 and CP400). The corresponding FeCl₃- and Fenton-modified biochars were also made using the above modification procedures (labeled as CFe300, CFe400, CFe-H₂O₂300 and CFe-H₂O₂400).

2.3. Characterization of rubber tree bark biochar

The textural property was analyzed using N₂ sorption/desorption isotherms at 77 K on a surface area and porosity analyzer (Micromeritics ASAP 2460, USA). The surface area was calculated by the Brunauer-Emmett-Teller (BET) method, and both pore volume and average pore size were calculated by Barret-Joyner-Halenda (BJH) method. The element content was analyzed with organic element analyzer (Thermo Scientific Flash 2000 CHNS/O, America) and inductive couple plasma (ICP) elemental analyzer (Agilent ICP-OES 730, America). The morphology was observed through scanning electron microscope (SEM, Phenom Prox, Holland) with energy-dispersive X-ray spectroscopy (EDS). The functional groups were characterized by Fourier transform and infrared (FTIR) spectrum (Bruker Tensor 27, Germany) using 2 mg biochar and 1000 mg KBr. The surface composition was characterized using X-ray photoelectron spectrum (XPS) (Thermo Scientific Escalab 250Xi, America). The crystallinity was determined on an X-ray diffractometer (Bruker D8 Advance, Germany) with Cu K_α radiation over a 2θ collection range of 10–90°.

2.4. Adsorption experiments

Briefly, biochar and adsorbate solution were added in conical flask and shaken at 180 rpm for desired time, then the mixture was filtered through a membrane filter for analyzing adsorbate concentration. For the kinetic experiments, the adsorption was carried out using the time intervals (0.5, 1, 2, 4, 8, 12, 16, 24, 32, 40 and 48 h). The adsorption isotherm experiments were performed using a series of initial concentrations (Cr: 100, 150, 200, 250, 300, 350, 400, 450, 500 mg/L; MB: 2.5, 5, 10, 15, 25, 50, 75 and 100 mg/L were applied for P400; 25, 50, 75, 100, 150, 200, 250, 300 and 350 mg/L were employed for Fe-H₂O₂400).

The MB and Cr (VI) concentrations were measured by UV–vis spectrophotometer (MAPADA UV-3300PC, Shanghai) at 665 nm and 540 nm [1.5-diphenylcarbazide was used as indicator for Cr (VI)], respectively. The adsorption capability of biochar was determined by the difference of concentrations before and after adsorption.

2.5. Regeneration of biochar

The regeneration of P300 and Fe-H₂O₂300 was performed in a pH13 NaOH solution (1/1 mg/mL ratio) for 16 h at 25 °C. The regeneration of PBC400 and FOBC400 was carried out in common solvents including CH₃CN, HCl solution and EtOH/acetic acid (9/1 vol ratio) using 1/2 (mg/mL) ratio for 24 h at 25 °C.

3. Results and discussion

3.1. Characterization

As shown in Table 1, both pristine and Fenton-modified biochars prepared at 300 and 400 °C possessed mesopore structure with inferior specific surface area (SSA). The Fenton-modified biochar had higher SSA and pore volume (PV) than the pristine biochar pyrolyzed at the same temperature.

The element analysis showed that Fenton-modified biochar

Table 1
Physical properties of biochar.

Biochar ^a	SSA (m ² /g)	PV (cm ³ /g)	PD (nm)
P300	1.11	0.00212	6.83
Fe-H ₂ O ₂ 300	2.63	0.01218	18.46
P400	2.11	0.00558	12.24
Fe-H ₂ O ₂ 400	3.47	0.00859	12.42

contained more Fe and O than corresponding pristine biochar (Table 2). Comparing to Fe-modified biochar, Fenton-modified biochar had much less Fe, implying H₂O₂ oxidation caused Fe release from biochar. It was noted that the O/C ratio of Fe-H₂O₂400 was much higher than that of P400 and Fe400, demonstrating organic oxygen containing groups could be generated on biochar matrix by Fenton oxidation.

The common functional groups including –OH (3448 cm⁻¹, stretching vibration), aliphatic C–H (2925 cm⁻¹, stretching vibration), C=O/aromatic C=C (1624 cm⁻¹, stretching vibrations), C=C (1435 cm⁻¹, stretching vibration), COOH (1383 cm⁻¹ bending vibration) (Song et al., 2014), aliphatic C–O (1058 cm⁻¹, stretching vibration) were observed on the pristine biochars prepared at 300 and 400 °C (E-supplementary data for this work can be found in e-version of this paper online). (Özçimen & Ersoy-Meriçboyu, 2010) Calcium oxalate (1317 and 781 cm⁻¹), (Tan et al., 2003) which might be derived from calcium salts on banana pseudostem, was also found on them. In comparison, the H₂O₂-modified biochars had weaker C–O stretching vibration peaks due to H₂O₂ oxidation. Obviously, the characteristic peaks of calcium oxalate were hardly observed on Fe-modified biochars, indicating the formation of calcium oxalate was completely inhibited during FeCl₃-promoted pyrolysis. Fe400 showed much stronger aromatic C–H bending peak (874 cm⁻¹) than P400, as aromatic structure was formed through dehydration of FeCl₃ at high temperature. As expected, new COOH (1710 or 1707 cm⁻¹, stretching vibration) and C–O (1058 or 1246 cm⁻¹, stretching vibration) groups were produced with Fenton modification. The C–O peak of Fenton-modified biochar shifted to higher wavenumber as pyrolyzed at relatively high temperature (400 °C) attributing to formation of aromatic structure. These suggested that Fenton oxidation surpassed single iron-promoted pyrolysis and H₂O₂ oxidation for generating oxygen containing groups on biochar.

The XPS analysis results were listed in Table 3. The Fenton-modified biochar had higher surface O content than corresponding pristine biochar. Iron was hardly detected on the surfaces of Fenton-modified biochar. Furthermore, the deconvolution of C 1 s peak showed that oxygen-containing groups including C–O (286.2 eV), C=O (287.2 eV) and COO (288.8 eV) (Yao et al., 2010) of Fenton-modified biochar were all more than that of corresponding pristine biochar. Therefore, various oxygen containing groups could be successfully introduced on the surface of biochar through Fenton oxidation.

SEM images demonstrated that pristine biochars were composed of irregular microparticles with rough surface (E-supplementary data for this work can be found in e-version of this paper online). Relatively smooth surfaces were obtained for Fenton-modified biochars comparing to pristine and Fe-modified biochars, indicating rough surface was damaged by Fenton oxidation. Obviously, a considerable amount of mesopores were generated on the microparticles of Fenton-modified biochars, thereby higher SSA and PV were obtained (Table 1). On the contrast, direct Fenton oxidation, where pristine biochar reacted with FeCl₃ and H₂O₂, rarely produced new mesopores even though H₂O₂ decomposition intensively occurred. Similarly, iron-promoted pyrolysis also hardly produced mesopores (E-supplementary data for this work can be found in e-version of this paper online). This indicated that the formation of porous structure mainly resulted from iron-promoted H₂O₂ decomposition taking place inside biochar matrix. In accordance with element analysis, EDS spectra also proved that the iron oxides and oxygen containing groups were incorporated onto pristine biochar with Fenton modification.

Pristine, Fe-modified and Fenton-modified biochars prepared at 300 and 400 °C were characterized by XRD spectra (E-supplementary data for this work can be found in e-version of this paper online). The multiple diffraction peaks at 2θ = 14.93, 24.39, 30.11, 38.18° were obtained for pristine biochars. These peaks were attributed to calcium oxalate hydrates (Kirboğa & Öner, 2009; Ouyang et al., 2004). Broad peaks around 2θ = 20.0° were also observed on them, indicating the presence of amorphous carbon (Xiong et al., 2018). Calcium oxalate

Table 2
Element analysis of biochar.

Sample	C (wt%)	H (wt%)	O (wt%)	N (wt%)	Fe (wt%)	O/C ^a	Fe/C ^a
P300	57.25	4.82	18.38	1.10	0.02	0.21	0.000
Fe300	52.63	4.64	31.57	1.08	5.21	0.40	0.021
Fe-H ₂ O ₂ 300	58.45	5.19	20.70	0.69	1.00	0.24	0.004
P400	62.09	3.78	20.85	0.86	0.07	0.22	0.000
Fe400	67.83	3.68	17.64	1.21	3.92	0.17	0.012
Fe-H ₂ O ₂ 400	57.45	3.48	30.93	0.62	1.68	0.36	0.006

^a mol ratio.

Table 3
Surface element and functional group composition.

Biochar	Element ratio				Functional group (binding energy, eV)			
	O/C	N/C	Ca/C	Si/C	C-C/ C=C (284.8)	C-O (286.2)	C=O (287.2)	O-C= O (288.8)
P300	29.84	0.00	0.12	3.44	1.00	0.20	0.03	0.12
Fe-H ₂ O ₂ 300	31.23	1.19	0.00	2.46	1.00	0.24	0.09	0.15
P400	24.09	1.26	1.99	0.00	1.00	0.18	0.03	0.09
Fe-H ₂ O ₂ 400	31.83	1.30	0.00	0.00	1.00	0.34	0.04	0.16

hydrates weren't produced with Fe-promoted pyrolysis as their feature diffraction peaks were hardly detected on the Fe-modified biochars, which was in consistency with FTIR analysis. Comparing to Fe300, additional diffraction peaks at $2\theta = 30.0^\circ, 35.3^\circ, 43.0^\circ, 56.8^\circ$ and 62.4° , which could be assigned to Fe₃O₄ (Duan et al., 2017) or γ -Fe₂O₃ (Zhang et al., 2013), were observed on Fe400. This indicated that the formation of magnetic iron oxide required high pyrolysis temperature (Han et al., 2016). Interestingly, subsequent Fenton oxidation generated crystalline graphene structure as a sharp peak of graphitic carbon at $2\theta = 26.60^\circ$ (JCPDS75-1621) (Lai et al., 2018; Zhi et al., 2014) was detected on Fenton-modified biochars. Moreover, it produced new magnetic iron oxide on Fe-H₂O₂400 as stronger diffraction peaks of Fe₃O₄ or γ -Fe₂O₃ were detected. Whereas, the magnetic iron oxides were scarcely generated on Fe-H₂O₂300. This suggested that the preformed magnetic iron oxide on biochar might act as an inducer for producing new one. More importantly, the saturation magnetization of Fe-H₂O₂400 was enhanced from 0.219 to 1.076 emu/g (Fig. 3c) with Fenton oxidation even though iron reduced (Table 2). It could be concluded that the magnetic iron of Fe-H₂O₂400 should be γ -Fe₂O₃ rather than Fe₃O₄ as low valence iron oxides including Fe₃O₄ could be oxidized into γ -Fe₂O₃ by H₂O₂. In a word, Fenton oxidation could be a promising method for improving the magnetic capability of magnetic biochar by converting nonmagnetic iron species into magnetic γ -Fe₂O₃.

3.2. Adsorption of Cr (VI) and MB

3.2.1. Effect of preparation parameters on adsorption capability

As shown in Fig. 1a, the optimal pyrolysis temperature was 300 °C for adsorption of Cr (VI) as Fe-H₂O₂300 displayed much higher adsorption capability than other pristine and Fenton-modified biochars. Different H₂O₂ concentrations (5, 10, 20 wt%) were found to be effective for improving the adsorption capability of Cr (VI) and 10 wt% H₂O₂ gave highest adsorption capability (Fig. 1b). For adsorption of MB, the optimized pyrolysis temperature and H₂O₂ concentration were 400 °C and 20 wt% H₂O₂, respectively (Fig. 1c-d). It was noted that the Fe-H₂O₂400 exhibited 9.8 times higher adsorption capability than P400. Thereby, Fenton modification is highly efficient for improving the adsorption capabilities for both Cr (VI) and MB. By contrast, iron-promoted pyrolysis only led to sharp decrease in adsorption of Cr (VI) and MB even incorporating iron oxides (Zhang et al., 2020). H₂O₂ post-oxidation caused significant diminishment and slight increase in

adsorption of Cr (VI) and MB, respectively. This demonstrated that Fenton modification outperformed single iron-modification and H₂O₂-modification. Besides, post-Fenton oxidation without iron-promoted pyrolysis failed to upgrade the adsorption capabilities of P300 and P400, thereby incorporation of Fe into biochar matrix through pyrolysis was of great importance for enhancing the adsorption capability of biochar. In the following experiments, the pristine and Fenton-modified biochars prepared at 300 and 400 °C were selected to study the adsorption behaviors of MB and Cr (VI), respectively.

3.2.2. The effect of initial pH on adsorption capability

The adsorption capabilities of P300 and Fe-H₂O₂300 toward Cr (VI) sharply decreased as increasing pH from 2 to 5 (Fig. 2a). This might be attributed to: 1) formation of HCrO₄⁻ and electropositive biochar surface, 2) increasing oxidizability of Cr (VI) at low pH (Zhou et al., 2016). For adsorption of MB, P400 always displayed inferior adsorption capability without impact of pH, and Fe-H₂O₂400 showed slightly increasing adsorption capability with the increase of pH from 3 to 9 (Fig. 2b). Additionally, the pH change of MB solution was recorded after adsorption to understand the adsorption process (Fig. 2c). The pH increased as performing the adsorption of P400 under acidic and nearly neutral conditions. Oppositely, the pH all decreased for Fe-H₂O₂400 over a wide pH range from 3 to 9. In particular, the pH sharply decreased to 3.8 while the adsorption was carried out at pH7. This indicated Fe-H₂O₂400 displayed strong cation exchange capability for adsorption of MB.

3.2.3. Adsorption kinetics and isotherms

The adsorption kinetics and isotherms of Cr (VI) and MB were depicted in Fig. 3. The adsorption kinetics were better described by pseudo-second order model (PSOM) than pseudo-first order model (PFOM) as PSOM offered good determinant coefficient and slightly higher calculated equilibrium adsorption capability than experimental one (Table 4). The adsorptions were all endothermic processes as the adsorption capabilities of biochars increased with temperature (Fig. 3e-h). The adsorption processes were better stimulated by Langmuir model than Freundlich model (Table 4). Fe-H₂O₂300 and Fe-H₂O₂400 exhibited much higher adsorption capacities than P300 and P400 with 335.57 and 258.14 mg/g maximum adsorption capabilities for Cr (VI) and MB at 45 °C, respectively, surpassed most biochars reported in the literature (Ma et al., 2014; Mian et al., 2018; Shi et al., 2018).

3.2.4. Regeneration of biochar

The regeneration of PBC300 and FOBC300 was performed in aqueous NaOH solution with pH13. Surprisingly, the adsorption capabilities of them increased for first regeneration (Fig. 4a), which could be resulted from the generation of new hydroxyls and carboxyls through hydrolysis of esters (Ding et al., 2016). As recycled for more than one time, both P300 and Fe-H₂O₂300 showed decreased adsorption capabilities probably owing to the incomplete desorption of Cr (VI).

The common solvents including CH₃CN, HCl solution and EtOH/acetic acid (9/1), which had been demonstrated to be efficient solvents for desorbing MB from carbon materials (Lyu et al., 2020), were used to regenerate P400 and Fe-H₂O₂400. However, only a small part of MB

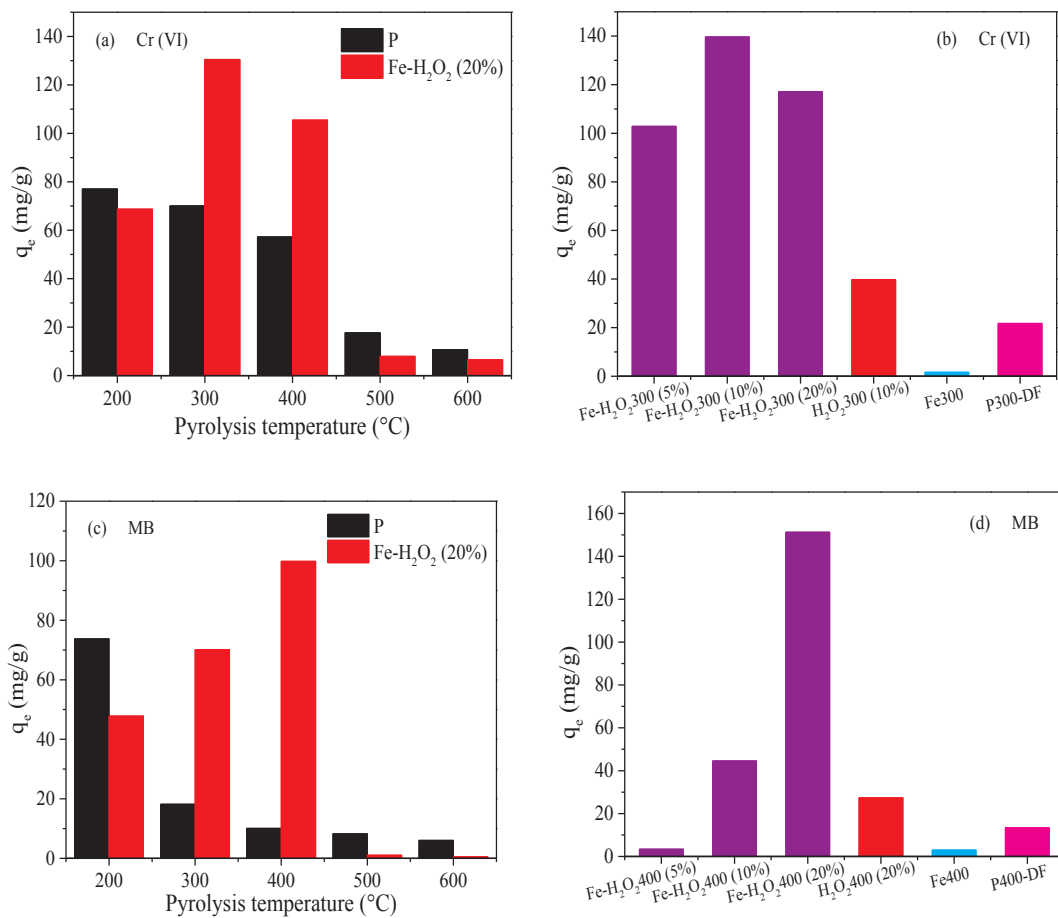


Fig. 1. Effect of preparation parameters on adsorption capabilities for Cr (VI) (200 mg/L, 1 g/L, pH2, 24 h (a) or 48 h (b), 25 °C) and MB (100 mg/L, pH7, 0.5 g/L; 24 h (c) or 48 h (d), 25 °C).

was desorbed, so the regenerated biochars showed much lower adsorption capabilities (Fig. 4b). This was also demonstrated by the FTIR spectrum of Fe-H₂O₂400 after desorption as the feature peaks of MB (1329, 1387 cm⁻¹) was still obviously present (E-supplementary data for this work can be found in e-version of this paper online).

3.2.5. Adsorption mechanism

The XPS analysis showed that the chromium loaded on P300 was mainly consisted of Cr (III) (Cr 2p_{3/2}: 577.3 eV, Cr 2p_{1/2}: 287.0 eV) (Liu et al., 2014; Manning et al., 2007) (E-supplementary data for this work can be found in e-version of this paper online). This indicated the removal of Cr (VI) by P300 was determined by reduction of Cr (VI) to Cr

(III). Differently, the chromium fixed on Fe-H₂O₂300 was comprised of Cr (III) (Cr 2p_{3/2}: 577.1 eV, Cr 2p_{1/2}: 286.7 eV) and Cr (VI) (Cr 2p_{3/2}: 579.0 eV, Cr 2p_{1/2}: 288.3 eV) with 67% and 33%, respectively (E-supplementary data for this work can be found in e-version of this paper online). Therefore, the removal of Cr (VI) by Fe-H₂O₂300 was attributed to both reduction of Cr (VI) to Cr (III) and sorption of Cr (VI). To gain deep insights into the adsorption mechanism, P300 and Fe-H₂O₂300 were examined by FTIR spectra after adsorption. The C-O peaks of the biochars (1100 and 1058 cm⁻¹) somewhat decreased (E-supplementary data for this work can be found in e-version of this paper online), suggesting C-O groups could act as reducing sites for removing Cr (VI). The stretching vibration peak of carboxyl (1710 cm⁻¹) on

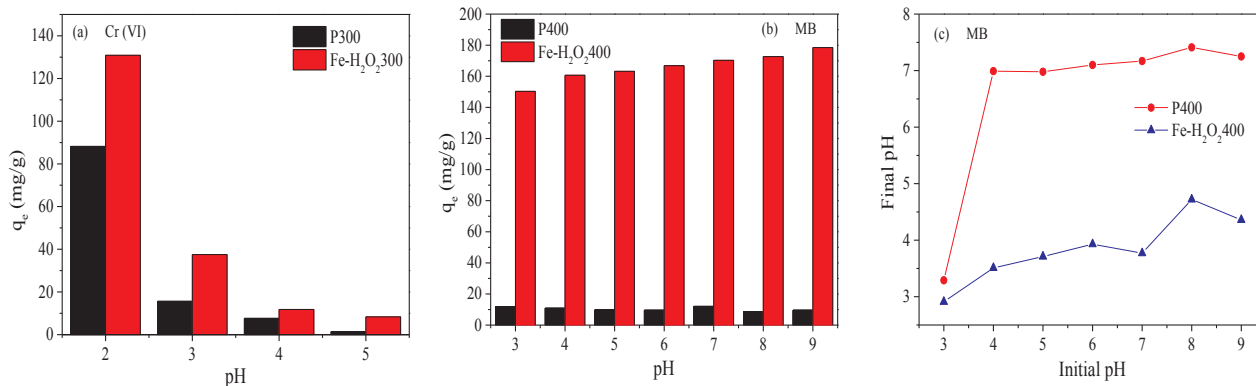


Fig. 2. Effect of initial pH on adsorption capabilities for Cr (VI) (200 mg/L, 1 g/L, 24 h, 25 °C) and MB (100 mg/L, 0.5 g/L, 48 h; 25 °C); and pH change of MB solution after adsorption (c).

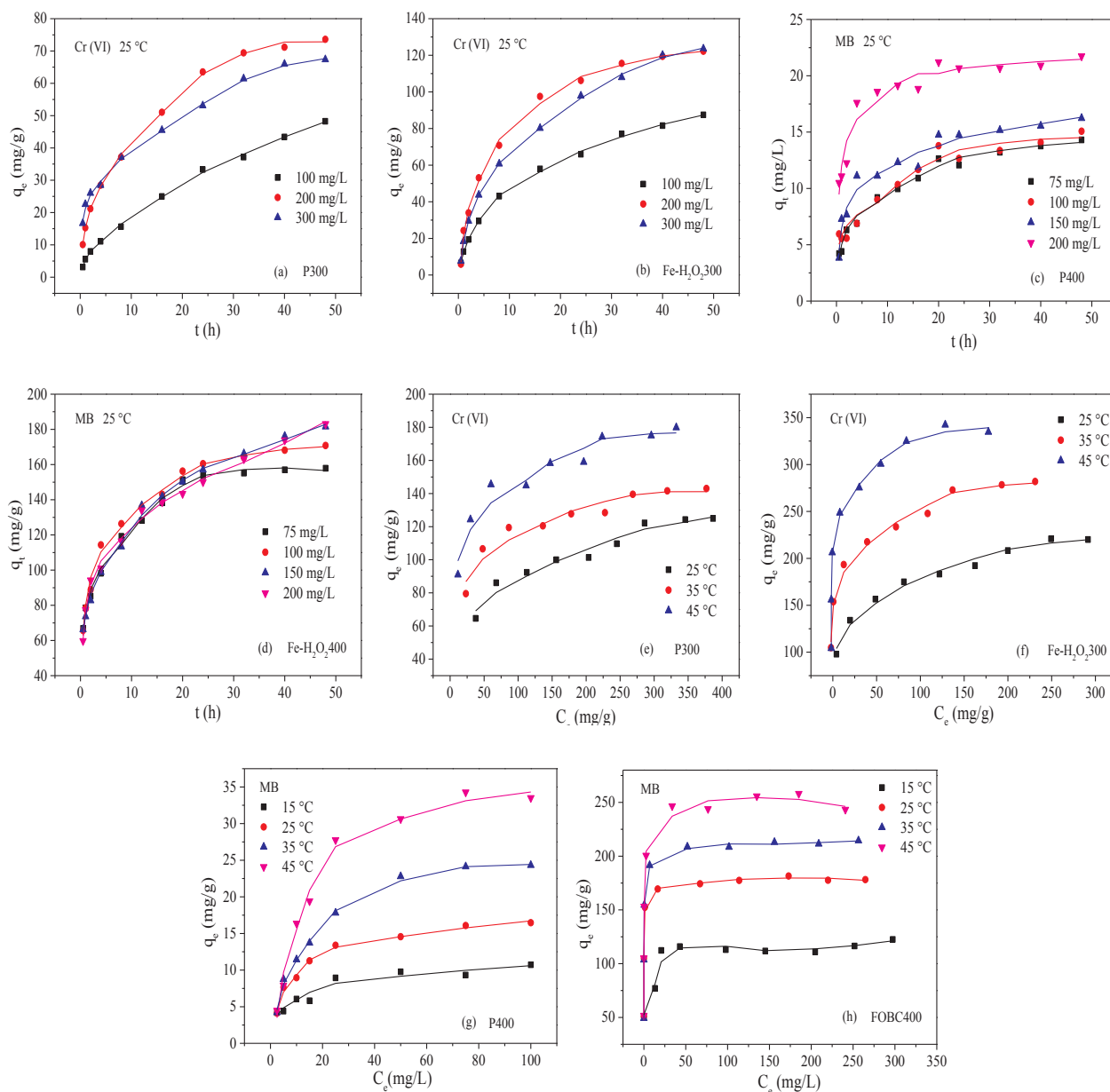


Fig. 3. Adsorption kinetics (a-d) and isotherms (e-h) of Cr (VI) (1 g/L, pH2) and MB (0.5 g/L, pH7).

FOBC300 completely disappeared probably ascribing to red shift and overlap by the peak at 1624 cm^{-1} (Fig. 9d). As Cr (VI) and Cr (III) mainly existed in the forms of HCrO_4^- and Cr^{3+} at pH2, respectively (Zhou et al., 2016), the red shift should be resulted from hydrogen bonding of HCrO_4^- and complexation of Cr^{3+} with carboxyl. Thus, hydrogen bonding and complexation could be involved in the Cr (VI) and Cr (III) adsorptions, respectively. Moreover, HCrO_4^- might be adsorbed above negatively charged surface at low pH and Cr^{3+} could be exchanged with proton of acidic groups such as COOH and phenolic OH.

Similarly, the FTIR spectra of P400 and Fe-H₂O₂400 were measured after adsorption to elucidate the adsorption mechanism of MB. Little change was observed for P400 due to less adsorption of MB (E-supplementary data for this work can be found in e-version of this paper online). Instead, red shift was observed for COOH peak of Fe-H₂O₂400 (from 1707 to 1701 cm^{-1}) and two feature C-N peaks (Gong et al., 2015) of MB (from 1398 to 1387 cm^{-1} and 1339 to 1329 cm^{-1}) (E-supplementary data for this work can be found in e-version of this paper

online). Due to partial release of MB, the COOH peak slightly shifted from 1701 to 1705 cm^{-1} after desorption. This demonstrated that hydrogen bonding was involved in the adsorption of Fe-H₂O₂400. pH obviously decreased after adsorption for Fe-H₂O₂400 as performing the adsorption at pH7 (Fig. 2c), indicating cation exchange between H^+ and MB cation occurred. Hence, carboxylic acids, which had strong cation exchange and hydrogen-bonding capabilities, should play an important role in the adsorption of MB. Additionally, cationic MB could also be adsorbed by electrostatic interaction of oxygen containing groups under natural conditions (Zhang et al., 2020). In brief, the generation of additional oxygen containing groups contributed to the increase of adsorption capability toward MB.

Iron oxides were less loaded on the surface of Fenton-modified biochars, contributed less to adsorption of Cr(VI) and MB. Therefore, the adsorption capabilities of Fenton-modified biochars were determined by biochar matrix. Considering only a small improvement in physical properties was obtained with Fenton modification (Table 1), the significant increase of adsorption capability toward Cr (VI) and MB

Table 4
The fitted parameters of adsorption kinetics and isotherms.

Kinetics	C ₀ (mg/g)	q _{e,exp} (mg/g)	PFOM: $\ln(q_e - q_t) = \ln q_e - k_1 t$			PSOM: $\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{1}{q_e}$				
			q _{e,cal}	k ₁	R ²	q _{e,cal}	k ₂	R ²		
Cr (VI)	P300	100	48.25	47.29	5.11×10^{-2}	0.9718	57.97	1.16×10^{-3}	0.9212	
		200	73.59	66.67	8.22×10^{-2}	0.9907	81.97	1.88×10^{-3}	0.9852	
		300	67.35	56.48	7.80×10^{-2}	0.9205	71.23	2.87×10^{-3}	0.9749	
	Fe-H ₂ O ₂ 300	100	87.48	79.20	6.27×10^{-2}	0.9896	99.30	1.12×10^{-3}	0.9863	
		200	122.20	108.71	8.87×10^{-2}	0.9926	141.44	9.48×10^{-4}	0.9896	
		300	123.59	120.90	7.59×10^{-2}	0.9404	143.27	7.51×10^{-4}	0.9870	
	MB	P400	75	14.31	9.86	7.13×10^{-2}	0.9744	14.93	1.72×10^{-2}	0.9893
			100	15.08	9.39	6.00×10^{-2}	0.8853	17.11	1.10×10^{-2}	0.9666
			150	16.24	9.32	6.96×10^{-2}	0.9286	16.38	1.86×10^{-2}	0.9728
		Fe-H ₂ O ₂ 400	200	21.75	7.42	6.97×10^{-2}	0.7238	22.53	2.46×10^{-2}	0.9810
75			157.88	95.93	1.17×10^{-1}	0.9824	166.67	2.88×10^{-3}	0.9972	
100			170.82	96.96	8.84×10^{-2}	0.9883	170.07	2.68×10^{-3}	0.9966	
150		181.54	117.63	7.04×10^{-2}	0.9795	187.27	1.57×10^{-3}	0.9897		
200		183.07	110.85	5.73×10^{-2}	0.9726	184.84	1.58×10^{-3}	0.9849		

Isotherms	T (°C)	q _{e,exp} (mg/g)	LM: $q_e = \frac{k_L q_{max} C_e}{1 + k_L C_e}$			FM: $q_e = K_F C_e^{1/n}$			
			q _{max}	K _L	R ²	K _F	1/n	R ²	
Cr (VI)	P300	25	124.97	140.85	0.01802	0.9867	25.83	0.2672	0.9548
		35	143.03	150.83	0.03870	0.9958	47.56	0.1901	0.9163
		45	179.82	185.87	0.05036	0.9940	62.61	0.1843	0.9346
	Fe-H ₂ O ₂ 300	25	220.79	229.89	0.05158	0.9908	74.80	0.1911	0.9950
		35	281.89	282.49	0.1676	0.9930	153.94	0.1065	0.9576
		45	342.28	335.57	0.9313	0.9962	196.23	0.1084	0.9458
MB	P400	15	10.72	10.62	0.2478	0.9876	4.86	0.1585	0.8332
		25	16.47	16.65	0.3482	0.9958	7.37	0.1851	0.9815
		35	24.32	25.01	0.3174	0.9946	10.22	0.1978	0.9688
	Fe-H ₂ O ₂ 400	45	34.28	34.00	0.7291	0.9972	17.43	0.1553	0.9876
		15	122.56	119.47	0.2239	0.9959	72.96	0.0905	0.8648
		25	178.18	178.90	2.5880	0.9998	114.98	0.0967	0.7708
	35	214.48	213.68	1.4904	0.9999	126.37	0.1155	0.7991	
	45	258.14	249.38	10.0214	0.9985	142.88	0.1281	0.8676	

should be primarily ascribed to introduction of new oxygen containing groups including -OH, C-O and -COOH.

3.2.6. Expansion of Fenton modification

When Fenton modification was applied to coconut shell biochar using 10 wt% H₂O₂, the adsorption capabilities of the biochars pyrolyzed at 300 and 400 °C were also greatly enhanced from 15.05 to 110.40 mg/g for Cr (VI) and from 6.81 to 205.05 mg/g for MB at 25 °C, respectively (Fig. 5a). The CFe-H₂O₂300 possessed inferior magnetic performance as the above Fe-H₂O₂300. Whereas, CFe-H₂O₂400 displayed 5.03 eum/g saturation magnetization, outperformed the corresponding Fe-modified biochar (CFe400: 3.77 eum/g) (Fig. 5b). This

further proved that Fenton modification was subjected to pyrolysis temperature for enhancing saturation magnetization of biochar. Importantly, a good saturation magnetization could be obtained with Fenton-modification, guaranteeing magnetic separation of biochar after adsorption.

4. Conclusion

A highly efficient strategy was established for improving the adsorptive and magnetic capabilities of biochar by combining FeCl₃-promoted pyrolysis and H₂O₂ oxidation (Fenton modification). The adsorption capabilities of rubber tree bark biochars toward Cr (VI) and

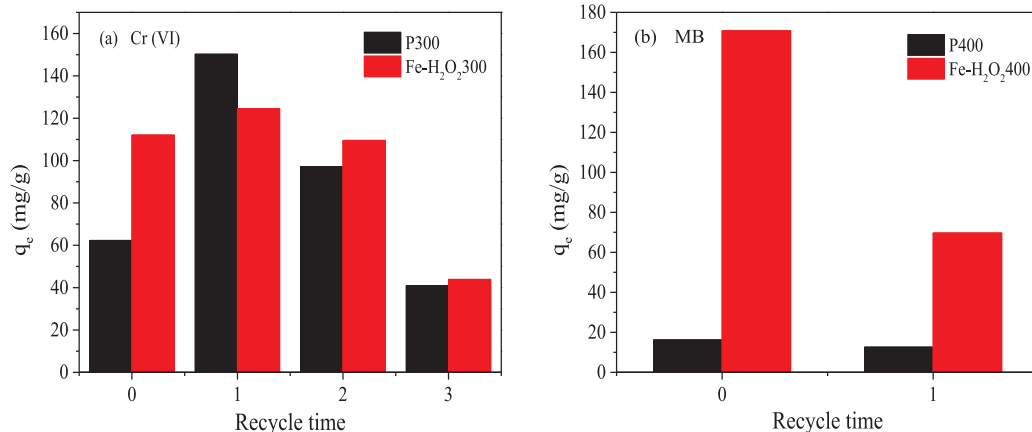


Fig. 4. The adsorption capability of regenerated biochar for Cr (VI) (200 mg/L, 1.0 g/L, pH2, 48 h, 25 °C) and MB (100 mg/L, 0.5 g/L, pH7, 24 h, 25 °C).

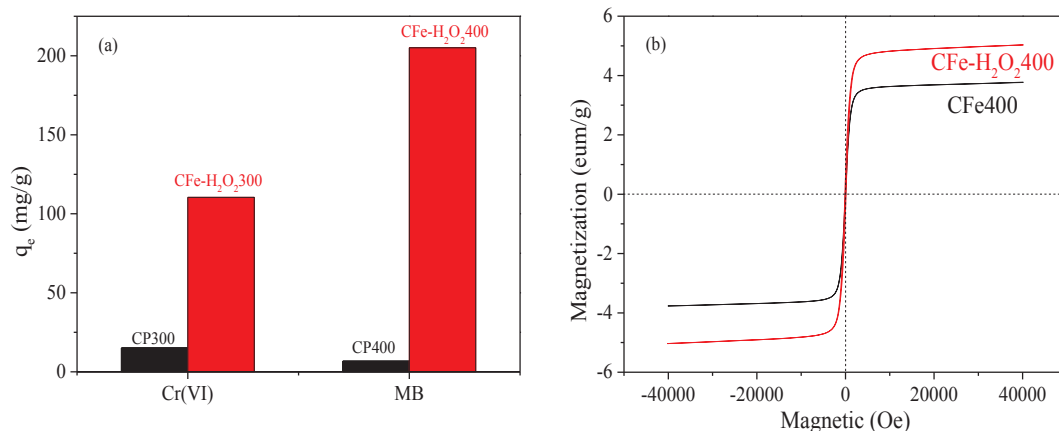


Fig. 5. The adsorption capabilities (a) (Cr (VI): 200 mg/L, 1 g/L, pH2, 48 h; MB: 200 mg/L, 0.5 g/L, pH7, 48 h) and magnetic hysteresis loops (b) of coconut shell biochar.

MB were enormously upgraded due to introduction of oxygen-containing groups. The saturation magnetization of magnetic biochar could also be enhanced by transforming nonmagnetic iron species into γ -Fe₂O₃. Moreover, this modification brought great enhancement in the adsorptive and magnetic capabilities of coconut shell biochar. This demonstrated that Fenton modification should be a general and efficient technology for improving the adsorptive and magnetic capabilities of biochar.

CRedit authorship contribution statement

Shuang Xu: Investigation, Data curation, Writing - original draft. **Jihui Li:** Conceptualization, Methodology, Writing - review & editing, Project administration. **Zhibing Yin:** Investigation, Visualization. **Sen Liu:** Investigation. **Siyao Bian:** Data curation. **Yucang Zhang:** Resources, Supervision.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.biortech.2020.123797>.

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