

Phenotypic Traits of Genetic Twins in Coconut Palm (*Cocos nucifera* L.) and their Utility in Seed Production

V. Arunachalam

ABSTRACT

Genetic identity is important in seed production of such sexually propagated, **cross-fertilized, heterozygous** crops as coconut palm (*Cocos nucifera* L.), which are **exclusively** propagated by seeds. Due to **their** uniform nature, **twins** are used in genetic studies, **breeding** and to determine identity. The objective was to identify uniform phenotypic traits withii twins **for use in determining** genetic identity. Phenotypic correlation was **determined among twin** pairs for 11 important **foliar** traits in **coconut**. **Of** the traits **under** study, leaf sheath fiber **thickness**, petiole **length** and number of leaflets had high correlation **and are** either highly heritable or have adaptive **significance**. These traits have practical applications in seed **production and breeding**.

INTRODUCTION

Seed purity and identity are important aspects of seed production and breeding for all crops. Coconut palm (*Cocos nucifera* L.) is an important **tropical** multi-purpose perennial crop. In the absence of vegetative propagation, the long pre-bearing age and the heterozygous nature of predominately cross-pollinated plants are impediments to breeding and propagation of wconut palm.

Twins have long been used in genetic studies due to their uniform nature (Christian et al., 1974). Twins are reported to **occur** in plants and have been used for multiplication, breeding (Bertrand and Cilas, 1990) and genetic identity studies (Cameron and Swra, 1973). However, the occurrence of twins or triplets is a rare phenomenon in wconut (Davis et al., 1953; Davis, 1979; Kumar and Rokhade, 1984). Polyembryony is a common feature in few varieties of mango (Aron et al., 1998) and citrus (Russo and Starrantino, 1975) controlled by single or multiple dominant **gene(s)**, respectively, but it only occurs in low frequency in **coconut** (Whitehead and Chapman, 1962).

Protein (Cardena et al., 1998) and molecular markers (Perera et al., 1999; Meerow et al., 2003) are used to analyze the genetic variation in coconut; however, identification of stable phenotypic traits would have great utility for breeding and selection. The cross sectional area of the petiole can provide an indirect estimate of the dry weight of the entire leaf (Friend and Corley, 1994). Leaf and petiole traits have also been shown to be important in diversity (Sugimura et al., 1997; Zizumbo and Garcia-Marin, 2001). The importance of leafsheath characters in classification (Nauman and Sanders, 1991), basic study (Tomlinson, 1964) and diversity of **coconut** palm (Arunachalam et al., 2005) have been reported by many-workers. Therefore, this study was undertaken to

evaluate 11 foliar traits in coconut twin pairs and their usefulness in identity determination.

MATERIALS AND METHODS

Seven twin pairs of 13 yr old coconut palms were used in the study. For each palm, observations were recorded on the oldest green leaf, which was removed from the stem. Petiole width, circumference and thickness were measured as described by Friend and Corley (1994). Thickness was measured with a vernier caliper.

The sheath from the youngest visible leaf on the crown of each palm was used in the study. Leaf sheaths were scored for presence or absence of dichotomous branching of the strands. If dichotomous branching was present for even a few strands, it was recorded as present. Three random fiber strands were separated from both warp and weh layers. A screw gauge was used to measure the thickness of these fiber strands. Correlation coefficient was determined between the pairs of twins for the foliar traits under study by use of chi-square tests ($P = 0.01$).

RESULTS AND DISCUSSION

Data on 11 foliar traits of seven coconut palm twin pairs is presented in Table 1. The phenotypic traits of leaf sheath fiber, number of Leaflets, length of petiole, cross sectional area of petiole and the leaflet number were highly correlated. However, the traits of petiole circumference and length of leaf bearing portion showed no relationship.

Most traits are similar among twins except for fruit weight, yield, female flower production, husk thickness and girth of nut (Krishna Marar and Kunhiraman, 1957). Louis (1981) observed that the number of leaves produced per year and the leaf length shows high genotypic variation in coconut and that these two traits are highly genetically advanced with strong potential

TABLE I. Comparison of 11 foliar traits in seven coconut palm twin pairs using chi-square tests.

No.	Trait	Observed	χ^2	$P \chi^2$
1	Number of leaves	0.82	0.04	0.99
2	Petiole circumference	0.08	10.58	0.1
3	Leaflet width	0.16	4.41	0.62
4	Petiole length	0.64	0.20	0.99
5	Length of leaf bearing portion	0.098	8.30	0.22
6	No. of leaflets	0.86	0.02	1
7	Leaf sheath fiber branching (+ or -)	0.98	0.000	1
8	Leaf sheath warp fiber	0.798	0.05	0.999
9	Leaf sheath weft fiber	0.69	0.14	0.999
10	Ratio of warp to weft fiber	0.93	0.005	1
11	Cross sectional area of petiole	0.25	2.25	0.895

$P = 0.01$

for **heritability** due to the predominance of additive genes. Of 19 morphological and physiological traits studied by Zizumbo and Garcia-Marin (2001), the number of leaves per unit time, leaf length and percentage of proximal rachis in leaf were reported to be discriminating traits **useful** in classifying five Mexican **ecotypes** of coconut.

Bourdeix (1999) utilized the number and width of coconut palm leaflets to describe the variation between **talls**, dwarfs and atypical dwarfs. He also noticed that dwarf **ecotypes** possess few leaflets and a smaller leaflet **surface area** than **tall** ecotypes. Niu Leka Dwarf, an **allogamous** (atypical) dwarf, had an equal number of leaflets to that of **talls**, but had the total surface area like dwarfs. The number of leaflets was also found to be one of the discriminating traits (Sugimura et al., 1997).

REFERENCES

- Aron, Y., H. Czosnek, S. Gazit, and C. Degani. 1998. Polyembryony in mango is controlled by a single dominant gene. *HortScience*. 33(7):1241–1242.
- Arunachalam, V., B. A. Jerard, V. Damodaran, M. J. Ratnambal, and P. M. Kumaran. 2005. Phenotypic diversity for **foliar** traits in Coconut germplasm. Genetic Resources and Crop Evolution: (In press).
- Bertrand, B. and C. G i . 1990. The use of true twins in experiments with cocoa. *L'utilisation de vrais jumeaux dans l' experimentation cacaoyere*. Cafe, Cacao-The. 34(4):295–298.
- Bourdeix, R. 1999. Selection and breeding. p. 117–195. *In* J. G. Ohler. 1999. Modern Coconut Management. Food Agric. Org. (FAO), Rome.
- Cameron, J. W. and R. W. Scora. 1973. Leaf oils and fruit characters in relation to genetic identity among twin and triplet citrus hybrids. *Lloydia*. 36(4):410–415.
- Cardeña, R., C. Oropeza and D. Zizumbo-Villarreal. 1998. Leaf proteins as markers useful in the genetic improvement of coconut palms. *Euphytica* 102:81–86.
- Christian, J. C., K. W. Kang and J. A. Norton. 1974. Choice of an estimate of genetic variance from twin data. *Am. J. Human Genetics*. 26(2):154–161.
- Davis, T. A. 1979. Some unusual formations in palms. *Principes* 23(2):80–83.
- Davis, T. A., A. P. Anandan and K. P. V. Menon. 1953. A case of polyembryony in coconut. *Indian Coconut J.* VI:67–71.
- Friend, D. and R. H. V. Corley. 1994. Measuring coconut palm dry matter production. *Exp. Agric.* 30:223–235.
- Krishna Marar, M. M. and C. A. Kunhiraman. 1957. Observations on a twin coconut palm. *Indian Coconut J.* X (2):3–8.
- Kumar, G. N. M. and A. K. Rokhade. 1984. Polyembryony in coconut. *Indian Coconut J.* 14(11):11.
- Louis, H. I. 1981. Genetic variability in coconut palm. *Madras Agric. J.* 68(9):588–593.
- Meerow, A. W., R. I. Wisser, J. S. Brown, D. N. Kuhn, R. J. Schnell and T. K. Broschat. 2003. Analysis of genetic diversity and population structure within Florida coconut germplasm using microsatellite DNA with special emphasis on the Fiji Dwarf cultivar. *Theor. Appl. Genet.* 106:715–726.
- Perera, L., J. R. Russell, J. Provan and W. Powell. 1999. Identification and characterisation of microsatellites in coconut (*Cocos nucifera* L.) and the analysis of coconut populations in Sri Lanka. *Mol. Ecol.* 8:344–346.

- Russo, F. and A. Starrantino. 1975. Studies on the heritability of *Polyembryony* in citrus. *Annali-dell'Istituto-Sperimentale-per l'Agricoltura*. 5:51-67.
- Sugimura, Y., M. Itano, C. D. Salud, K. Otsuji and H. Yamaguchi. 1997. Biometric analysis of diversity of coconut palm: *cultivar* classification by botanical and agronomic *traits*. *Euphytica*. 98:29-35.
- Tomlinson, P. B. 1964. The vascular skeleton of coconut leaf base. *Phytomorphology* 14(2):218-230.
- Whitehead, R.A. and G. P. Chapman. 1962. Twinning and *haploidy* in *Cocos nucifera*. *Nature*. 195:1228-1229.
- Zizumbo-Villareal, D. and P. GarciaMarin. 2001. Morpho-physiological variation and phenotypic plasticity in Mexican populations of coconut. *Genetic Res. and Crop Evol.* 48(6):547-554.