

Isolation of microbial pathogens of *Aceria guerreronis*, the coconut eriophyid mite

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Abstract

A multi-disciplinary team consisting of microbiologists, pathologists, entomologists and nematologists for the isolation of possible pathogens, specifically *Hirsutella thompsonii*, from the coconut eriophyid mite, *Aceria guerreronis*, conducted a collaborative study during the month of May 2001, when the mite population was at its peak in Kerala. One hundred and sixty mite-infested coconuts were collected from Thrissur District for this purpose. Isolation of microbes from these nuts was done by plating the mite-wash and its serial dilutions, surface-sterilizing the whole nut and then plating the mites and the perianth tissue and plating of the individual mites. It was observed that the actinomycetes constituted the predominant microflora (seven isolates) during this period followed by yeasts (four isolates); fungi (three isolates) and bacteria (two isolates). Pathogenicity trials in the laboratory revealed that fungi caused mortality to the mites in the range of 10-20%; actinomycetes, 15-20% and bacteria, less than 10%. Yeasts were non-pathogenic to the mites. The well-known mite pathogen *Hirsutella thompsonii* was not encountered/isolated during this study.

Key words: *Aceria guerreronis* (eriophyid mite), coconut, *Hirsutella thompsonii*, microbial pathogens

Introduction

Microbes are one of the most diverse groups of organisms inhabiting the earth. This diversity makes them an ideal choice to work with for improvement of agriculture. One of the important areas where microbes make an impact is the ecologically safe and specific management of pests. However, this involves obtaining virulent pathogen(s) for which extensive isolations and screening of a wide variety of the microorganisms is necessary (Gopal and Gupta, 2001). A sound understanding of the diversity and population dynamics of the microbes associated with the host and its habitat, becomes paramount for such studies. The eriophyid mite, *Aceria guerreronis*, has become an important pest of coconut since its observation in 1998. It has been reported

that microorganisms, especially the fungus *Hirsutella thompsonii*, is the long-term solution for management of this acarine pest. In order to isolate possible pathogens associated with this mite specifically *H. thompsonii*, a rapid-fire and multi-disciplinary effort involving microbiologists, pathologists, entomologists and nematologists was undertaken during the month of May 2001, when the mite population was at its peak in Kerala. Results of this effort are outlined in this research paper. Communication.

Materials and Methods

Around 160 mite-infested nuts were collected from the KAU experimental farm, Vellanikkara and other farms of Thrissur District of Kerala. The nuts were mostly 3-9 months age with varying degrees of mite damage.

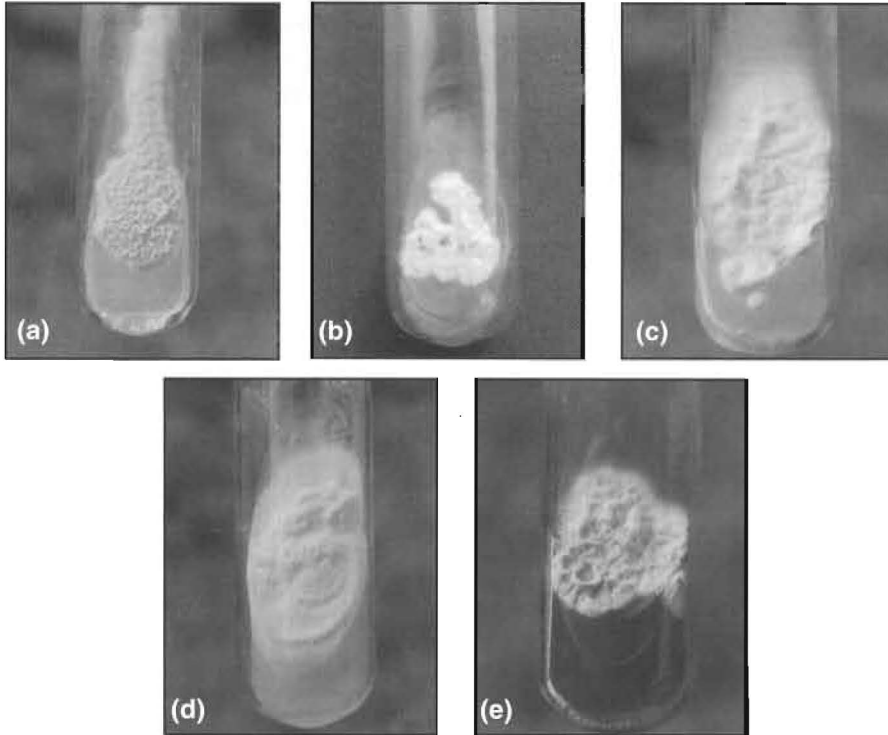
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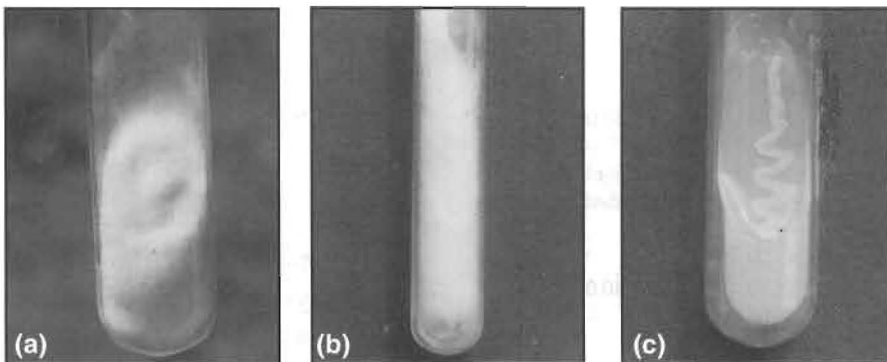
28-30 were brought to the laboratory and grouped in such way that each group had nuts representative of the different areas from where they were collected. These different nuts were distributed to the different discipline teachers. Isolation of the microorganisms was done by following methods : (a) plating of mite washings as (b) by serial dilution of mite washings, (c) surface sterilizing the whole nut and then plating the mites and mite tissue and (d) plating of individual mites. Different agar, potato dextrose agar and Kuster's agar

media were used for isolation of different groups of microorganisms (bacteria, fungi and actinomycetes). The plates were incubated at 30 °C and the resultant colonies were purified. Pathogenicity bioassays with the purified microbes were conducted on the mite colonies by spraying their cell suspensions, grown in their respective growth media, on the detached nuts. The control nuts were applied with sterilized broth only. The mortality of the mites was recorded by washing technique, 5 days after the application of the treatments.



1. Actinomycetes isolates giving more than 17.5% mortality of mite

- EM Act 2 (isolated from Kodagara, Thrissur from an unattended coconut garden)
- EM Act 4 (isolated from Aloor, Thrissur from an organically maintained coconut garden)
- EM Act 5 (isolated from Aloor)
- EM Act 6 (isolated from Instructional Farm plot, KAU, Thrissur)
- EM Act 7 (isolated from Vattakad, Thrissur, from a garden which was sprayed with neem oil-garlic mixture)



1. Fungi and bacteria isolated from eriophyid mite

- EM Fungi (*Fusarium* sp isolated from mite and perianth tissue of coconut from instructional Farm plot, KAU, Thrissur)
- EM Fungi (*Fusarium* sp with pinkish pigmentation, giving pathogenicity of 18.5%, isolated from Kodagara, Thrissur)
- EM Bacterium (unidentified, isolated from CPCRI, Kannara, Thrissur)

Results and Discussion

Bucher (1960) reported that screening of the micro flora associated with the insects and their significance may lead to isolation of possible pathogens. Isolations made from the coconut eriophyid mite during the peak infestation period i.e. April to May (Anon., 1999) revealed that actinomycetes were the predominant micro flora with 7 types of isolates obtained with an isolation frequency of 54%. Yeasts were the second dominant group of microbes with 4 types of isolates and an isolation frequency of 40%. The fungi could be isolated at a frequency slightly higher than 30%, and *Aspergillus*, *Penicillium* and *Fusarium* were the commonest fungi observed. The bacterial isolates were obtained at the least frequency (10- 20%) and *Serratia marcescens* was one of the bacteria identified from the isolations. Detailed results are presented in Table-1. The succession of the micro flora association was not acutely dependent on the infestation stages of the mite. In the early stages of the mite colony development, rarely any microbial association was observed. With the advancement in infestation, yeasts were the earliest to be isolated. As the infestation reached the middle stage with browning of the nuts visible externally, fungi, actinomycetes and bacteria started appearing during the isolations.

Table 1 Types and frequency of microorganisms isolated from coconut eriophyid mite during its peak infestation season (May, 2001)

Type of microorganism	Frequency of individual isolation (%)	Average frequency (%)
Actinomycetes		
EM*-Act1	45.00	54.00
EM-Act2	50.00	
EM-Act3	70.00	
EM-Act4	30.00	
EM-Act5	55.00	
EM-Act6	60.00	
EM-Act7	70.00	
Fungi		
<i>Fusarium</i> sp	30.00	32.00
<i>Aspergillus</i> sp	45.00	
<i>Penicillium</i> sp	20.00	
Yeasts		
EM-Yeast1	30.00	40.00
EM-Yeast2	45.00	
EM-Yeast3	35.00	
EM-Yeast4	50.00	
Bacteria		
<i>Serratia marcescens</i>	20.00	17.50
Unidentified bacterium	15.00	

* EM stands for eriophyid mite

It was noted that while plating the mites, perianth portion or nut tissue, the actinomycetes and fungi were growing in abundance. However, when washings of the mites were plated, *Serratia marcescens* dominated in the plates (Table 2). Yeasts were seen to grow with ease in both the plating techniques. Our observations are supportive of earlier reports of isolation of *Fusarium* and actinomycetes from coconut eriophyid mite (Hall et al., 1980). *Aspergillus* sp. and *Penicillium insectivorum* have been reported to be pathogenic to cassava green mite *Mononychellus tanajoa* (Bartowski et al., 1988) and *Ixodius ricinus* (Kalsbeek et al., 1995). Isolation of bacteria - *Pseudomonas* and *Bacillus*, yeasts and actinomycetes were also reported in our earlier studies (Gopal et al., 2002), but *Serratia marcescens* has been observed to be associated with the mites for the first time. Another important observation is the preponderance of the actinomycetes association with the mite during the summer period and when the mite infestation is at its peak. Actinomycetes are known to flourish in high temperature and low moisture habitats. These conditions could be prevailing in the nut microenvironment during the summers. However, there is enough moisture to sustain some of the common genera of fungi that were reported in this study. Notwithstanding the actinomycete domination during the summer months as reported in this paper, fungi are the overall preponderant microorganisms associated with this mite when observed throughout the year (Gopal et al., 2002).

Table 2 Growth of microorganisms as per the isolation technique employed

Isolation technique	Microorganism
Simple mite plating	Actinomycetes, Fungi and yeasts
Plating mite from surface sterilized nut	Fungi, Actinomycetes and yeasts
Plating of perianth pieces with mite colonies	Fungi, actinomycetes, yeasts
Plating of perianth pieces with mite colonies from surface sterilized nut	Actinomycetes, fungi, yeasts and bacteria
Plating of mite colony wash	<i>S. marcescens</i> , Unidentified bacteria, yeasts
Plating of mite colony wash after serial dilution	<i>S. marcescens</i> , Unidentified bacteria, yeasts
Plating of nut tissue with mite colony	Actinomycetes, fungi, yeasts and bacteria
Streaking of mite(s) on agar plates	Actinomycetes, fungi, yeasts and bacteria

Mortality of the mites in the pathogenicity bioassays conducted with the actinomycetes, bacteria and yeasts is outlined in Table 3. The actinomycetes gave a maximum mortality of 15% followed by the fungi with 10 to 20%. *S. marcescens* gave a maximum of 10% mortality of the 4 yeasts were found to be non-pathogenic.

organism to be designated as a potential pathogen, it is necessary that it kills at least 50% of the target host. In this study, a maximum of 25% death of the mites could be achieved by the application of actinomycetes and the application of the microbes gave still lesser mortality. The conclusion is that besides the fungus *Scopulariopsis* and *Chaetium* which gave 27% mortality of coconut eriophyid mite (Gopal *et al.*, 2002), the actinomycetes also bring about the same as seen in this study.

Table 1. Results of pathogenicity assay with isolated microbes on detached eriophyid mite affected coconuts

Microorganism	Mortality of mites (%)
Act1	17.5
Act2	23.5
Act3	14.5
Act4	25.0
Act5	22.0
Act6	18.5
Act7	23.5
Chaetium sp	18.5
Streptomyces sp	15.0
Chaetium sp	9.0
Yeast1	2.0
Yeast2	2.5
Yeast3	0.0
Yeast4	1.5
<i>Aceria marcescens</i>	7.5
Identified bacterium	8.5
Control broth	5.0
Dextrose broth	7.5
Water's broth	7.5
Control broth	2.5

Results are an average of three replications with 5 mites in each replication

The difference in the extent of mortality caused by different category of the microbes is attributed to the mode of action and their preponderance. The actinomycetes are reported to produce toxins like streptolydigin, melbectimycin and spinosad, which have acaricidal/acaricidal properties. The bacterium *S.*

marcescens kills the insects only on ingestion, however they also produce proteolytic enzymes capable of killing insects. Fungi as mentioned earlier are best suited for mite suppression.

Isolation of the well-known acarine pathogenic fungus *Hirsutella thompsonii* from the coconut mite collected from Thrissur area had been reported by Beevi *et al.*, (1999), and from Pollachi by Sreerama Kumar and Singh (2000). However, it was not encountered/isolated in this study or during our earlier work (Gopal *et al.*, 2002). More thorough and regular isolation studies carried out through different weather periods and on coconuts collected from different mite-infested locations will definitely yield potential pathogen(s) for the long-term management of this important coconut pest.

References

- Anonymous. 1999. Annual Report, *Central Plantation Crops Research Institute*, 88 p.
- Bartowski, J., Odindo, M.O. and Otieno, W.A. 1988. Some fungal pathogens of the cassava green spider mites, *Mononychellus* spp. (Tetranychidae) in Kenya. *Insect Science and its Application* **9**: 457-459.
- Beevi, Pathummal, S., Beena, S., Lyla, K.R., Varma, Sukumara, A., Mathew, P. Maicykutty and Nadarajan, L. 1999. *Hirsutella thompsonii* var. *synnematosata* Samson, McCoy and O'Donnell on coconut mite *Aceria (Eriophyes) guerreronis* Keifer a new report from India *J. Tropical Agric.*, **37**: 91-93.
- Bucher 1960. Potential bacterial pathogens of insects and their characteristics. *J. Invertebrate Pathology* **2**: 172-195.
- Gopal, Murali and Gupta, Alka. 2001. Has *Hirsutella thompsonii* the wherewithal to counter the coconut eriophyid mite scourge. *Current Science*, **80**: 831-836.
- Gopal, Murali., Saleena, A. and McCoy, C.W. 2002. Study of microflora associated with coconut eriophyid mite as a preliminary step towards pathogen isolation. *Current Science* **82** (1): 22-24.
- Hall, R.A., Hussey, N.W. and Mariau, D. 1980. Results of a survey of biological control agents of the coconut mite *Eriophyes guerreronis*. *Oleagineux* **35**: 395-398.
- Kalsbeek, V., Frandsen, F. and Steenberg, T. 1995. Entomopathogenic fungi associated with *Ixodes ricinus* ticks. *Acarology*, **19** : 45-51.
- Sreerama Kumar, P. and Singh, S.P. 2000. *Hirsutella thompsonii* : the best biological control option for the management of the coconut mite in India. *Indian Coconut Journal* **31**(5): 11-17.